An-Najah National University Faculty of Graduate Studies

# Assessment of TiO<sub>2</sub> as Photocatalyest for Complete Mineralization of Aqueous Bacteria and their Organic Content

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This Thesis is Submitted in Partial Fulfillment of the Requirements for the Degree of Master of Chemistry, Faculty of Graduate Studies, An-Najah National University, Nablus - Palestine.

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## **Dedication**

My heart pulsates with the thrill for tendering gratitude to those persons who helped in completion for my project. To my mother, father, wife, daughter, son, sister and brother for their continuous support with my appreciation.

The pleasant point of presenting a report is the opportunity to thank those who have contributed to build my knowledge. Unfortunately, the list of expressions of thank no matter how extensive is always incomplete and inadequate. Indeed, this page of acknowledgement shall never be able to touch the horizon of generosity of those who tendered their help to me.

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v الإقرار

أنا الموقع أدناه مقدم الرسالة التي تحمل العنوان:

## Assessment of TiO<sub>2</sub> as Photocatalyest for Complete Mineralization of Aqueous Bacteria and their Organic Content

أقر بأن ما اشتملت عليه هذه الرسالة إنما هو نتاج جهدي الخاص، باستثناء ما تمت الإشارة إليه حيثما ورد، وأن هذه الرسالة ككل أو جزء منها لم يقدم من قبل لنيل أي درجة أو بحث علمي أو بحثي لدى أي مؤسسة تعليمية او بحثية أخرى.

## Declaration

The work provided in this thesis, unless otherwise referenced is the researcher's own work, and has not been submitted elsewhere for any other degree of qualification.

**Student's Name:** 

Signature:

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Date:

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	light condition.				

Abbreviation	Term
a.u.	Arbitrary unit
ANU	An-Najah National University
AOPs	Advanced oxidation processes
CO <sub>2</sub>	Carbon dioxide
СВ	Conduction band
eV	Electron volt
Eg	Energy band gap
$\lambda_{\text{EX}}$	Excitation wavelength
$\mathbf{h}^+$	Holes
OH•	Hydroxyl radical
Μ	Molarity
SEM	Scanning Electron Microscopy
TiO <sub>2</sub>	Titanium dioxide
UV-Vis	Ultraviolet-Visible
VB	Valence band
XRD	X-Ray Diffraction
WHO	World health organization
UV	Ultraviolet light
TOC	Total Organic Carbon
S. aureus	Staphylococcus aureus
PL	PhotoLuminescence Spectra
NPs	Nano- particles
G	Gram
e	Electron
UAE	United Arab Emirates University
QY	quantum yield
CFU	colony forming units
TN	Turnover Number

## xi List of Abbreviations

## xii Assessment of TiO<sub>2</sub> as photocatalyest for Complete Mineralization of Aqueous Bacteria and their Organic Contents

By Raed Rasmi Hassan Ali Supervisor Dr. Ahed Zyoud Co-Supervisor Prof. Hikmat Hilal Abstract

Photodegradation is one of the most useful methods of water purification because it reduces pollution levels by eliminating harmful bacteria in the water and includes at a later stage the mineralization of chemical pollutants or pollutants within the water. Photo catalytic activity using titanium dioxide  $TiO_2$  utilizes the longest wavelength located at the end of ultraviolet. In this work water was purified from positive bacteria by complete mineralization using of both types  $TiO_2$  (Rutile) and  $TiO_2$ (Anatase) using UV tail in solar simulated radiation. The results obtained revealed the high efficiency of  $TiO_2$  (Anatase) in the destruction of bacteria and mineralization. This catalyst was able to destroy the bacteria G +ve S. aureus and baptized after exposure to 4 hours of light. The catalyst also proved its ability to inhibit bacteria in the dark but to a lesser extent than under radiation. It is therefore recommended to expand the present study and include other types of biological pollutant.

# Chapter One Introduction

1

## **1.1 General**

Water is the condition of life. It occupies seventy one percent of earth surface. Only about (2.5%) of earth waters are fresh, round most of this is found in polar areas (~99%),while in rivers, lakes and the atmosphere 0.3% of fresh water is present [1, 2].

Recently, fresh water challenge accrued in many parts of the world, UN statistics estimated that about one billion people on the planet [3] have no effective means of disinfection of water [4]. Non-potable water resulting after human usage is called gray water, which can be treated with ease [5].

Black water is a wastewater that requires comprehensive treatment. water pollution occurs on an individual and industrial level, especially in developing countries [6]. Low water quality has harmful effect especially on human health and the environment in general. The World Health Organization (WHO) estimates that about 1.4 million children die annually by diarrhea from contaminated water [7].

Environmental Pollution is one of the dangerous challenges to the human as it negatively affects daily lives. Environmental pollution includes Water pollution due to waste from factories and houses [8]. Air pollution caused respiratory diseases by sulfur and nitrogen oxides [9]. Purification may itself cause other problems. Therefore, natural resources must not be harmed from the beginning [10].

However, as contamination is a de facto, we must look for alternatives to address this problem by finding new products that do not cause additional damage to the environment while working to restore the environmental conditions using natural energy sources such as sunlight [11]. Solar light is a renewable low cost nonhazardous energy.

### **1.1.1 Biological pollution of the aquatic environment**

Biological pollution refers to the presence of organisms in the fresh or saline water. Pathogenic microorganism, such as bacteria, viruses, parasites, algae and protozoa , also contaminates water [12, 13].

Pathogens are spared from human and animal waste directly to fresh or salt water bodies, or indirectly through sewage or agricultural water. Such contaminants cause different diseases. Therefor contaminated water should be treated. Examples for treatment are chlorination, ozonation, mechanical filtering and others [4].

Water may contains safe species such as (oocytes, larvae, infectious phases) [14], and some parasites like schistosomiasis, hepatic worms and insets like mosquitoes.

#### **1.1.2 Biological contaminants**

Living organisms in water may interact with the aquatic environment, leading to imbalance in the environments, known as biological pollution [15]. Some of these organisms are more widespread than others in certain environments due to the nature and size of those organisms [16] and the

spread or scarcity of natural enemies in addition to competition between organisms in the same environment. For example, one gram of agricultural soil may contain more than 200 million bacterial cells and 300 thousand mushroom Parasites [17].

## 1.1.3 Bacteria

Bacteria are the most prevalent microorganisms found in animal and human feces. Drinking stool-contaminated water is the major reason of waterborne infections, along with some bacteria. Fortunately only a few cells in water are pathogenic [18].

### **1.1.4 G+ve and G-ve bacteria**

Bacteria are two types based on the ability to color with the Gram staining technique (Gram's negative bacteria and Gram's positive bacteria) [19]. Gram-positive bacteria (G+ve) bacteria maintain the color of the dye and have bacterial cell wall with thick peptidoglycan surface [19]. (Crystal violet to expands in Gram positive bacteria and yields purple gram pigment, making it need iodine to keep the crystal violet trapped inside the layer Peptidoglycan [20]. (Table1) Gram-negative bacteria (G-ve) contain a

3

thin cell wall of peptidoglycan, which is trapped between the cell membrane and the external bacterial membrane. There is additional protection from proteins and saccharide grease, which makes the bacteria resistant against antibodies due to the impenetrable cell wall [20]. They cause wide range of therapeutic cases (Table1).

	Gram negative bacteria	Gram positive bacteria
Gram reaction	Can be decolourized to accept counter stain (Safranin or Fuchsine) stain red or pink , they don't retain the G stain when washed with absolute alcohol and acetone.	Retain crystal violet day and stain dark violet or purple, they remain colored blue or purple with G stain when washed with absolute alcohol and water.
Peptidoglycan layer	Think	Thick
	(single layered)	(multilayered)
Outer membrane	Present	Absent
Resistance to physical disruption	Low	High
Cell wall composition	The cell wall is 70-120 A Think : two layered . Lipid content is 20-30% (high), Murein content is 10-20% (low).	The cell wall is 100-120 A Think : single layered. Lipid content of the cell. Wall is low , whereas Murein Content is 70- 80% (higher).

Table 1.1: Comparison between G-ve and G+ve bacteria [20-22].

## 1.1.5 Staphylococcus aureus

These are Gram positive (G+ve) types with non-moving small round shape.

They are approximately 0.5-1.5 µm in diameter [23, 24].



Figure 1.1: G+ve stain S.aureus cell.

Although it does not cause disease all the time, one of the diseases caused by this type of bacterium is the toxic shock syndrome, which leads to a severe disease accompanied by fever, widespread red rash with other organs affected [25, 26]. In environment S. aureus is relatively common but is mainly found on the skin and mucous membranes of animals. In the gastrointestinal track. S. aureus is rarely found and can be detected within sewage [24, 27]. Human contact with water environment like swimming pools, spa pools and other recreational waters may release S. aureus. It was also found in domestic water well [28].

## **1.1.6 Water purification**

Water disinfection means the elimination, deactivation or mineralization of pathogenic microorganisms. Microorganisms are destroyed or deactivated resulting in termination of growth and reproduction. Decontamination is a process related to disinfection [29]. However, during decontamination

process all present microorganisms are killed, both harmful and harmless microorganisms [30, 31]. Disinfection can be chemical or physical. For physical disinfection of water several disinfectants can be utilized such as ultraviolet light (UV), electronic radiation, gamma rays and heat [32]. For chemical disinfection of water several disinfectants can be used such as chlorine dioxide (ClO<sub>2</sub>), Ozone (O<sub>3</sub>). Hypochlorite ion (OCl<sup>-</sup>), halogens including chlorine (Cl<sub>2</sub>), bromine (Br<sub>2</sub>) and iodine (I<sub>2</sub>), metal ions such as copper (Cu) and silver (Ag), potassium permanganate (KMnO<sub>4</sub>), alcohols, soaps and detergents [31].

	Advantages		Linitations	
Chlorine(Cl <sub>2</sub> )	Very effective	Well-	Chlorine is	
	for removing	established	dangerous gas	
	almost all	technology	that is lethal at	
	microbial	more cost-	concentrations	
	pathogens	effective	as low as 0.1	
		than either	percemt air by	
		UV	volume	
Chlorine	Very effective	radiation or		
Dioxide Clo <sub>2</sub>	even in low	ozone		
	concentration;	disinfection	Denserated	
	Droad-	; chiorine	should be	
	auick and long	nrolong	stored in a	
	time	disinfection	dark area	
	disinfection:	: reliable	uarkarca	
	dose not	and		
	generate	effective		All forms of chlorine are highly
	chlorinated	against a		corroive and toxic ; create
	phenols and	wide		hazardous compounds-
	THMs	spectrum of		disinfection by – products
Sodium	Easier to	pathogenic	Very corrosive;	(e.g.,trihalomethanes(THMs))
Hypochlorite	handle ; no	organisms	decompose and	
Solution	disinfection by	effective in	should not be	
	products	oxidizing	stored for more	
		certain	than one	
		inorganic	month; stored	
		compounds	dry area	
Solid calcium	When	; flexible	ury area.	
Hypochlorite	packaged	dosing		
, F	calcium	control;		
	hypochlorite is	eliminate		
	very stable	certain		
	allowing long	noxious		
	time storage.	odors		
 Chlanamina			A	
Unioramine	An enective		A COFFOSIVE	
	produces fewer		strong odor	
	disinfection hv-		that requires	
	products:		proper hndling	
	generated		: should be	
	onsite;		kept away	
	Chloramine		from organic	
	forming		materials;	
	reactions are		readily absorbs	
	99 percent		moisture,	
	complete		forming	
	withina few		chlorine gas.	
0	minutes.			
Ozonation	Requiring shorte	r contacttime	Uzone gas is unst	able and must be generated onsie
	and dosage than	emorine ;	; does not mainta	in an adequate residual in water
	destroying viruse		, requiring comp	ncated equipmet and entrient
	ucsu oying vii use	6		

 Table 1.2: Comparison between main disinfectants [35-[33].

## How does disinfection work?

Disinfection commonly takes place by damaging the cell wall of bacteria, demoralize proteins or nucleic acids synthesis, changing cell membrane permittivity and effecting enzymes action. These dis-arrangements in cell activity inhibit microorganisms to grow [34, 35]. Oxidizing disinfectants also remove organic matter from water, and remove nutrients Figure (1.2) shows a sketch for the photo-catalytic inactivation of a bacterial cell process [36, 37].



Figure 1.2: Inactivation of a bacterial cell [36, 37].

#### **Disinfection by oxidation processes.**

Advanced Oxidation Processes (AOP) are among the modern chemical techniques used for water disinfection [38]. Abiotic degradation (such as wet oxidation, chemical oxidation or acid-base hydrolysis) and photo-degradation by  $H_2O_2/UV,O_3/UV$  or  $O_3/H_2O_2/UV$  processes, solar photolysis, processes in vacuum ultraviolet or photo-catalysis are studied [34, 39].

#### What does Photo -catalysis Mean?

The word photo-catalysis is two parts. Part one "photo" means "light" and part two "catalysis" means "stimulation "of the catalysis process by a material. This material is considered a catalyst. It increases the rate of reaction by decreasing the energy needed to activate reaction [40]. Photo catalysis is therefore a process of light activation to increase the rate of chemical reaction without having role in the reaction itself. Photo-catalyst functions and uses can be divided into basic categories.. Water purification, Prevention of contamination, antibacterial, removing unpleasant odors, air purification [41, 42]. All of these applications in the presence of photocatalyst depend on sunlight or ultraviolet light from any source [43].

Semiconductors have been chosen as photo-catalysts because they have relatively small energy gaps between the valance band and the conduction band [44, 45] as shown in Table (1.3).

semiconductor	Eg(ev)
Si	1.1
Fe <sub>2</sub> O <sub>3</sub>	2.3
TiO <sub>2</sub> (rutile)	2.5
TiO <sub>2</sub> (anatase)	3.0
ZnO	3.2
SnO <sub>2</sub>	(3.2-3.7)

 Table 1.3: Band gap width for common semiconductor [46, 47].

In photo catalysis, the semiconductor absorbs energy from the rays of the sun, or from an ultraviolet source that is at least equal to the energy gap [48].

A positive hole result and can oxidize molecules. In photo-catalysis, radiation excites the catalyst and the reactants are oxidized by oxygen as shown in Figure (1.3).



Figure 1.3: The role of titanium dioxide in photo-catalytic degradation [49, 50].

## 1.1.7 Titanium dioxide

Among semiconductors that can be used as a photo-catalysts is titanium dioxide, with the chemical symbol  $TiO_2$  and energy gap Eg=3.2 Ev [51, 52].



Figure 1.4: Titanium dioxide powder TiO<sub>2</sub>.

Titanium dioxide is best suited for several advantages as a photo-catalyst.

For example it is inert, corrosion resistant and needs less refining and preparation than other semiconductors, It is available at a low price under normal conditions.

Titanium dioxide absorbs ultraviolet light with enough energy to create electron and a positive hole. The electron in the titanium dioxide becomes excited and the electron  $e^-$  travels to the conduction band leaving a positive hole (h <sup>+</sup>) valence band. In this case, dioxide of titanium becomes excited.

The titanium dioxide hole h<sup>+</sup> turns the water molecule into hydrogen and hydroxyl radical. The electron e<sup>-</sup> reacts with dissolved oxygen and creates a very strong oxidizing anion  $O_2^-$ . This process goes on with radiation [53].



Figure 1.5: The mechanism of semiconductor photo-catalyst [54][55].

The reaction mechanisms are widely known and can be understood by Equations [55-57].

$$TiO_2 + h\upsilon \rightarrow e_{cb}^{-} + hv_b^{+}$$
<sup>(1)</sup>

$$h_{vb} + H_2O \rightarrow H^+ + HO^{\bullet}$$
<sup>(2)</sup>

$$h_{vb+} + R \rightarrow \text{intermediates} \rightarrow CO2 + H2O$$
 (3)

$$\mathbf{e_{cb}}^{-} + \mathbf{O}_2 \to \mathbf{O}_2^{+-} \tag{4}$$

$$HO' + R \rightarrow intermediates \rightarrow CO_2 + H_2O$$
 (5)

It is generally known that the photo-catalytic disinfection of water requires exposure to irradiation of equivalent or higher energy than the band gap of the semiconductor. Zinc Oxide is classified as a semiconductor in group II- VI, whose covalence is on the boundary between ionic and covalent semiconductors. A broad energy band (3.2-3.37 eV) with high thermal and mechanical stability at room temperature makes it attractive for potential uses [25].

Titanium dioxide(TiO<sub>2</sub>), has unique physical and chemical properties, such as high chemical stability, high electrochemical coupling coefficient, broad range of radiation absorption and high photo-stability [58].

Titanium dioxide (TiO<sub>2</sub>), has emerged as an excellent photo-catalyst material for environmental purification and it used in the area of photo catalysis, mainly photo catalytic air purification, sterilization and cancer medication [48].

#### **1.2 Literature review**

In 2015, various morphologies of ZnO Nano-materials were synthesized and used as photo-catalyst for self-sensitized photo-degradation of Malachite Green (MG) under solar light ZnO flowers exhibited highest photo-catalytic activity, with complete mineralization of dye to produce  $CO_2$  [59].

In 2016, Venieri and his team worked on doping titania with non-metals to decrease its band gap energy to less than value of 3.2 eV. Disinfection effect of cation-doped titania in S. aureus elimination in aqueous samples under artificial and natural sunlight was investigated. For Fe, Al and Cr doped TiO<sub>2</sub>, the relative activity for S. aureus inactivation decreased in the

order Fe-TiO<sub>2</sub>> Al-TiO<sub>2</sub>> Cr-TiO<sub>2</sub> with 99.9% deactivation of S. aureus population after 60 min [60].

In 2017, a novel combined solar system with a  $TiO_2$  thin-film photo reactor and a pasteurizer operating under continuous flow was tested. both bacteria inactivation and organics degradation where studied [61]. In 2017, electrochemical oxidative degradation of diazo dye amido black 10B (AB10B) as model pollutant in water has been studied by using nanostructured ZnO-TiO<sub>2</sub> thin films deposited on graphite felt (GrF) substrate as anode [62].

In 2009 Gurudev Sujatha and his team studied the applicability of UV photo catalysis for the treatment of coffee processing wastewater with  $TiO_2$  as a catalyst. The experimental runs were carried out with, wastewater produced by an Indian company. In addition to the applicability of the photo-catalysis, parameters were tested, namely, first wastewater pH, load of catalyst, UV light potential, kind of catalyst and oxidant loading. Finally, the use of solar light instead of UV was tested , and the performances obtained in both cases were compare the use of photo-catalysis [56].

### **1.3 Objectives**

The main purpose of this work is "Disinfection of water by complete deactivation and mineralization of G+ve bacteria by using a low cost and safe process of photo mineralization".

## **1.4 Novelty of this work**

Literature showed that  $TiO_2$  nano-particles can inhibit bacteria under solar light However literature does not describe what happens to the inhibited bacteria. As bacteria involve organic matter, which can be hazardous to humans, such matter may remain in water. Literature did not describe how to remove such organic materials from drinking water.

This proposed work is a novel. It will the assess ability of  $TiO_2$  photocatalysts to completely inhibit bacteria and mineralize their organic content under direct solar light. Such study has not been described in literature to our knowledge.

## Chapter Two Materials and Methods

## **2.1 Materials**

## 2.1.1 Titanium dioxide (TiO<sub>2</sub>)

Two types of titanium dioxide granules (Anatase and Rutile) were used. Commercial powders (catalog no 22,422-7 and 1317-80-2) were taken from (Sigma Co.) and used in photo-degradation of contaminated water samples.

## 2.1.2 Bacteria

Positive Gram+ bacteria named S.aureus was used. This type of bacteria was isolated from clinical sample obtained from Department of Medical Science laboratory at an-Najah National University.

## 2.1.3 Other chemicals

Ethanol, sodium hydroxide and hydrochloric acid were purchased from Aldrich –sigma Co. or from Frutarom Co.

Nutrient broth was purchased from Hi Media Lab Pvt. Ltd, India and nutrient agar was bought from USA company Sparks, Becton, Dickinson.

#### **2.2 Instruments**

#### **2.2.1 UV–visible spectrophotometry**

A Labo Med, Inc. 1601spectrophotometer was used to measure bacterial concentration. Quantitative determination of the bacterial concentration was made through the use of tubative method. By measuring the amount of light that passed through bacterial contaminated water. The more suspended bacterial cells show more turbidity, and more turbidity means less light passing through the suspension.

The wavelength 630 nm was used in the absorption range (0.08-0.1) where the initial concentration of bacteria equals  $1.5 \times 10^8$  CFU\mL. This is consistent with the standard McFarland turbidity.

#### **2.2.2 Photoluminescence spectra device (PL)**

Perkin-Elmer LS50 Luminescence Spectrophotometer was used to measure the emission fluorescence spectra for  $TiO_2(0.1g)$  in 50 ml water anatase and rutile. Excitation wavelength was (360) nm.

The emitted spectrum was used to calculate the semiconductor catalyst band gap. The gap was compared with the literature [46, 47, 63-65] where the spectrum was studied for each of commercial anatase and commercial rutile.

#### 2.2.3 Light sun simulator

An intermediate type sunlight simulator lamp is used. Maximum 400 watts. The intensity of the radiation passing through the water was controlled in order to obtain accurate result.

## 2.2.4 Lux-meter

A Lux-meter (Lx-102 light meter) was used to measure the intensity of light or radiation pass through tested polluted water to determinate the efficiency of naked catalyst. and was set to ~100000 lux (similar to solar light intensity about 1,360 watts per square meter).

#### 2.2.5 XRD

X-ray diffraction was used to identify and examine crystallization of the catalyst. The patterns for dry powder were recorded using Philips XRD XPERT PRO diffractometer with CuK with X-ray wavelength ( $\lambda$ =1.45 Å). The analysis was kindly performed at UAE University, Al Ain, UAE.

### 2.2.6 SEM

SEM photos of the samples were taken using a Jeol Model JSM-6700F. The analysis was performed at UAE University, Al Ain, UAE.

#### 2.2.7 TOC

The Total Organic Carbon (TOC) was used as an equivalent to the organic content of water in order to assess the degree of mineralization of treated aqueous solution. Inorganic carbon was determined by the acidification of the sample in the rate 1% hydrochloric acid was measured before injection and organic carbon was burned ~at 800 C°. The total Organic Carbon (TOC) was calculated using a SHIMADZU Model TOC-L CSH\CSN. Each measurement was repeated three times, and the average was taken for nearest values. The analysis was kindly performed by Faculty of Agricultural Science and Technology at Palestine Technical University.

#### 2.3 Preparations

#### **2.3.1** Preparations related to bacterial treatment.

G+ve S.aureus bacteria were used. Nutrient agar were prepared according to manufacturer's instructions and poured after sterilized in dishes in a regular manner in order to facilitate the counting process The nutrient broth was prepared according to the manufacturers. Instruction and then poured into the test tube 5ml of each tube sterilize by autoclave.

Saline solution was prepared in concentration of (0.9 %) by dissolving 9.00 g of NaCl salt in  $1000_{mL}$  distilled pure water and then poured into test tube at rate of 9  $_{mL}$  each tube then sterilized by autoclave. The previously prepared saline solution is used to prepare fold dilutions of bacteria in polluted water samples to count bacteria.

All prepared materials and tools were autoclaved at 121 °C under 1.5 atm, and kept in retreater (~4°C) for the sake of different analysis.

In order to prepare the bacteria we take , a few bacterial colonies were taken and put them in 5 mL nutrient broth tube and incubated for period of 2-4 hours at 37 °C [66]. Then the sample was analyzed by using UV–Visible spectrophotometry at 630 nm wavelength and then bacteria were diluted until device reading was in the range (0.08 - 0.10) [67]. It expresses the concentration of McFarland were concentration of bacteria is  $1.5 \times 10^8$  CFU/mL [68, 69].

In all experiments  $1 \times 10^6$  CFU/mL bacterial concentration was treated. This concentration was obtained by the following equation :

(Molarity 
$$_1 \times Vol_1$$
) con = (Molarity  $_2 \times Vol_2$ ) dil.

$$(1.5 \times 10^{8} CFU/mL_{1} \times \text{Vol}_{1})_{\text{con}} = (1 \times 10^{6} CFU/mL_{2} \times 50mL_{2}) \text{ dil.}$$

Vol <sub>1</sub> taken from concentrated bacteria solution = 0.330 mL.

Where, Moarity<sub>1</sub> is McFarland concentration which obtained by UV– Visible spectrophotometry at 630 nm wavelength ( $1.5 \times 10^8$  CFU/mL).

Vol <sub>1</sub> is taken from McFarland concentration. Molarity <sub>2</sub> is needed bacterial concentration to be treated in 50 mL sample. Vol<sub>2</sub> is sample volume (50 mL).

#### 2.4 Catalytic experiment

#### 2.4.1 Bacterial disinfections

As shown in Fig. (3.1), each container includes 50 mL of distilled and sterile water. The first was called a control sample that contained only bacteria. The second container contained bacteria treated with  $TiO_2$  anatase. where both pots were exposed to sun simulated light in specific and multiple period of time and two other pots were made each containing 50 ml of sterile water the first cup contains bacteria only, the other contains bacteria treated with (TiO<sub>2</sub>) anatase where placed in the dark under same condition and in specific and multiple time period.



Experimental steps for disinfection of bacteria

Figure 2.1: Experimental steps for bacterial deactivation by using several catalyst systems [70].

Finally, for all experiments and at the end of selected time,  $1000 \ \mu$ L of each treated sample was withdrawn using a micropipette and diluted in a series of saline solution tubes with different dilutions (0.1, 0.01, 0.001 dilutions) [71].

From each diluted, aliquots of 0.1 mL were taken by yellow tip and were spread on two nutrient agar Petri dishes using bent glass rod, then incubated for 24 hours at 37 °C. After incubation, the bacteria were grown and colonies in each plate were counted using plate count method, only plates which have colonies between 30 and 300 colonies were considered.

The average number of each two plates for each dilution was calculated and the bacteria concentration as CFU/mL unit was reported according to the following equation:

Bacterial conc = (Average number of colonies  $\times$  Dilution factor  $\times 10$ ) [72].

The % deactivation = (Conc. initial – Conc. final )  $\div$  Conc.intial×100%)

- Conc.final is bacterial final concentration.
- Conc.initial is bacterial initial concentration.

where the initial concentration was taken from samples contained bacteria only under sacrifice condition (control sample), and the final concentration was taken from samples treated with  $TiO_2$ . Experiments were tested on a G+ve .S.aureus bacteria.

#### **2.4.2 Impact of various parameters on bacteria mineralization**

The effects of various parameters on photo-mineralization efficiency were studied as follows:

## 2.4.2.1 Time effect

Different reaction times (10, 30 and 60 min) were tested. Experiments are made with constant (0.1g) catalyst amount and bacterial concentration  $1 \times 10^6$  CFU\mL at temperature 28 °C and (pH~ 7) both TiO<sub>2</sub> anatase and rutile were used against S.aureus and under irradiation using sun light simulator. After 10 minutes the differences were observed in ability to destroy bacteria using TiO<sub>2</sub> anatase and TiO<sub>2</sub> rutile system. After 60 minutes total bacterial deactivation was achieved

#### 2.4.2.2 Influence of bacteria concentration

Effect of concentration of bacteria  $(1 \times 10^6, 3 \times 10^6, 5 \times 10^6 \text{ CFU}\text{mL})$  on the mineralization efficiency of S.aureus studied [73].

#### 2.4.2.3 Effect of pH

Based on earlier literature PH values (5.7, 7.3, and 8.08) these pH values are suitable for bacteria growth were used to study the effect on the mineralization of bacteria loss [34, 74-76].

## **2.4.2.4 Temperature effect**

Bacteria can live at (20 °C,30 °C, and 37 °C) these temperatures used to study the effect of temperature on the mineralization of bacteria [25, 67, 77].

## 2.4.2.5 Effect of catalyst amount

Catalyst quantities (0.1,0.05 and 0.025g) TiO2 anatase were used for economical purposes. A constant concentration of bacteria (1×106) CFU/mL at 28 °C and pH =7.3 under the sun–simulation irradiation was used. measurements were made after (10, 30, and 60 min).

## 2.4.2.6 Catalyst reuse

After the completion of the mineralization process, the treated solution was completely sterilized. Then it was contaminated with a quantity of bacteria  $(1 \times 10^6)$  CFU/mL. The process was repeated again twice in order to study the catalyst, recovery and reuse.
# Chapter Three Results and Discussion

#### **3.1 Preliminary remarks**

The main aim of this research was to disinfect water by complete deactivation and mineralization of a G+ve (S. aureus) bacteria by using a low price, secure method passed on with solar light. Naked TiO2 anatase and rutile were characterized using PL, Electronic Absorption Spectra, SEM, and XRD instruments.

TiO<sub>2</sub> systems were then used to catalyze the photo-mineralization of G+ve

(S. aureus) using simulated light. Complete mineralization of bacteria and their organic contents was tested by measuring TOC in reaction mixture. Finally, factors impacting photo-mineralization reaction and catalyst efficiency were studied.

#### **3.2 Catalyst characterization**

## **3.2.1** Photoluminescence spectra (PL)

The photoluminescence (PL) emission spectra were measured for commercial (TiO<sub>2</sub> anatase) and (TiO<sub>2</sub> rutile). The purpose is to find  $E_g$  for each TiO<sub>2</sub> system. As shown in Fig.(3.1), PL spectra collected using 300 nm excitation wavelength show maximum wave lengths.



PhotoLuminescence (PL) spectra for TiO2 both system

**Fig 3.1:** (PL) spectra of 0.1 g  $\text{TiO}_2$  dispersed in 50 mL distilled water under 300 nm UV excitation wavelength, for commercial anatase (A) and rutile (R) system.

By using equation ( $E_g$  (eV)= h×c ÷  $\lambda$  max (nm)) or (eV=1.2398÷  $\lambda\mu_m$  max) [78, 79] were h is blank constant ,c is light speed in space and  $\lambda$  is maximum wavelength recorded and intense emission peaks shown in Fig.(4.1).

The E<sub>g</sub> values for TiO<sub>2</sub> systems were calculated using (Eg (eV)=  $h \times c \div \lambda$  max (nm)) or (eV=1.2398÷  $\lambda\mu$ m max), as brief in Table(4.1).

Table 3.1: Wavelengths (nm) found on highest point in PL spectra and energy Band Gap determined by Eg equation, for TiO<sub>2</sub> systems.

	Commercial TiO <sub>2</sub> anatase	Commercial TiO <sub>2</sub> rutile
λ(nm)	415	425
Eg(eV)	2.987	2.91

As in Table (4.1) TiO<sub>2</sub> systems give (2.987, 2.91 eV) band gap values. Our results are within the literature range of TiO<sub>2</sub> (2.98-3.05 eV). Eg gives indication about particle size.

When Eg value increases, particle size decreases Since the small particle has low band gap energy and thus shifted to the absorption to the lower wavelength, somehow, a higher particle size behaves contrarily [80-82].

## **3.2.2 SEM results**

Both commercial  $TiO_2$  (anatase and rutile ) were studied using Scanning Electron Microscopy.



Figure 3.2.a: SEM images for solid TiO<sub>2</sub> powders commercial anatase.



Figure 3.2.b: SEM images for solid TiO<sub>2</sub> powders commercial rutile.

The micrographs show that the commercial rutile  $TiO_2$  has large agglomerates (~230 nm), involving nanoparticles (average ~50 nm) as confirmed by XRD below. Based on SEM, the anatase  $TiO_2$  has agglomerates (~250 nm), with average nanoparticles in the range 45-50 nm as confirmed by XRD( smaller particle), as shown below.

## 3.2.3 XRD results

The average crystallite sizes of the samples were studied by using Debye Scherer's equation DP= $0.94\lambda$ \ $\beta$ cos $\theta$  [83], using the full width at half maximum height of the X-ray diffraction peaks, Where, Dp = Average Crystallite size,  $\beta$  = Line broadening in radians,  $\theta$  = Bragg angle,  $\lambda$  = X-Ray wavelength.



Figure 3.3: XRD patterns for commercial (TiO<sub>2</sub> anatase systems).



## 2Theta (degree)

Figure 3.4: XRD patterns for commercial (TiO<sub>2</sub> rutile systems).

Table (3.2) summarizes Crystallite size in D (nm) for both commercial TiO<sub>2</sub> catalysts, based on Debye Scherrer's equation [84-88].

Table 3.2: Average crystallite size of commercial  $TiO_2$  anatase and rutile calculated by XRD.

X-ray diffraction peaks	Crystallite size, D(nm) for commercial TiO <sub>2</sub> anatase	X-ray peaks	diffraction	Crystallitesize,D(nm)forcommercialTiO2
101	41.53		110	rutile
200	41.55		110	53.01
211	47.76		101	51.15
	<b>Average = 45.13</b>			<b>Average = 50.72</b>

Table (3.2) shows that average crystallite size for commercial  $TiO_2$  anatase catalyst was (45.13 nm) and for commercial  $TiO_2$  rutile catalyst was (50.72 nm). Smaller size for anatase means higher surface area of catalyst which increases its efficacy in disinfection of polluted water.

By these results we expect that anatase catalyst will be more efficient than rutile commercial catalyst in deactivating bacteria.

Both SEM and XRD results summarize the particle size for both  $TiO_2$  systems. Both SEM and XRD results show that anatase powder has smaller crystallite size than commercial rutile.

## **3.3 Simulated solar irradiation experiments**

Both  $TiO_2$  anatase and  $TiO_2$  rutile catalysts were used against G+ve S. aureus bacteria under solar simulated radiations. Performances of  $TiO_2$ anatase catalysts were found with quantum yield (QY), turn over frequency (TF) and deactivation percent, which are simply calculated by following methods: QY = bacterial colonies concentration lost experimentally/number of incident photons (n)

Energy (J) = Incident power per unit area  $\times$  Exposure time (s)  $\times$  exposed area (cm<sup>2</sup>).

Assuming average wavelength of incident light is 630 nm, then  $\upsilon=c/\lambda$ , and E(J) = nh $\upsilon$ 

Where:

M. wt of  $TiO_2 = 79.866$  g/mol.

Incident power per unit area for solar light =  $0.0146 \text{ w/cm}^2$ 

exposed area = 23.75 cm<sup>2</sup>, c =  $3 \times 10^{8}$ , h =  $6.62 \times 10^{-34}$ 

Average wavelength of incident light for UV light = 630 nm, n = number of photon.

Turnover Number (TN) = Number of deactivated bacteria / unite of TiO<sub>2</sub> [89], TF = TN / Time.

deactivation percent = [Number of deactivated bacteria / Number of initial bacteria] $\times 100\%$ .

Incident power per unit area for solar light =  $0.0146 \text{ w/cm}^2$  [90].

## 3.3.1 Impact of time on bacterial deactivation

Fig. (3.5) illustrates the influence of exposure time on the deactivation percentage of  $(1 \times 10^6 \text{ CFU}\text{mL})$  of G +ve S. aureus bacteria treated by 0.1 g of (anatase and rutile TiO<sub>2</sub>) in separate experiments under light.



**Figure 3.5.a:** Influence of exposure time on the deactivation percentage of  $(1 \times 10^6 \text{ CFU} \text{mL}) \text{ S}$ . aureus bacteria, catalyst loading 0.1g ( anatase TiO<sub>2</sub> catalyst) at 28 °C under light condition and pH=7.4



**Figure 3.5.b:** Influence of exposure time on the deactivation percentage of  $(1 \times 10^6 \text{ CFU}\text{mL})$  S. aureus bacteria, catalyst loading 0.1g (rutile TiO<sub>2</sub> catalyst) at 28 °C under light condition and pH=7.4

Fig. (3.5.a) and Fig.(3.5.b) shows that after 60 min 70.29% (S.aureus bacteria) were deactivated when we use anatase TiO<sub>2</sub>. only 24% was deactivated when we use rutile TiO<sub>2</sub>. Therefore, shorter exposure time (10 min) and (30 min) was necessary to compare the effectiveness of (anatase TiO<sub>2</sub>) and (rutile TiO<sub>2</sub>). The deactivation percent by anatase TiO<sub>2</sub> which was 48.27% after 30 mint compared to Rutile TiO<sub>2</sub> which was (only 19%) for S. aureus treatment.

The deactivation percent of anatase  $TiO_2$  was to 22.11% after 10 mint compared to Rutile  $TiO_2$  (only 10% after 10 min) for S. aureus treatment.

We conclude from this result that (rutile TiO<sub>2</sub>) is less effective than (TiO<sub>2</sub> anatase) in the process of deactivate bacteria. Therefore anatase will be used in the subsequent processes due to its high efficiency to deactivate this type of bacteria. Tables (4.4) give summery values of quantum yield (QY), turnover frequency (TF) and deactivation percent of  $(1 \times 10^6 \text{ CFU}/\text{mL})$  of S. aureus bacteria using 0.1 g of TiO<sub>2</sub> anatase in light for (10, 30 and 60) min.

Table 3.3: Values of QY, TF and deactivation percent for different exposure times. All experiments were done using 0.1 g of TiO<sub>2</sub> anatase against S.aureus ( $1 \times 10^6$  CFU\mL) at 28 °C and light condition.

Time	QY	<b>TF×10<sup>-17</sup></b> min <sup>-1</sup>	TN×10 <sup>-16</sup>	% bacterial deactivation
10 m	1.377×10 <sup>-38</sup>	1.327	1.372	22.2%
30 m	1.28×10 <sup>-44</sup>	1.239	3.712	48.27%
60 m	1.06×10 <sup>-44</sup>	1.169	9.416	70.29%

Table (3.3) shows that OY and TF values. Were higher at the penning of treatment, due to higher bacterial concentrations, which promoted more deactivation for both bacteria types. As the reaction begins, bacterial

concentration was lowered which caused decrease in the deactivation percent per time unit [91, 92]. Another reason is also possible. As reaction progressed, organic molecules resulting from deactivated bacteria become more and catalyst functioned to decompose both these organic molecules and the remaining bacterial cells [25, 93]. Fig. (3.6.a) and (3.6.b) shows the influence of exposure time on the deactivation percentage of  $(1 \times 10^6$ CFU\mL) of G+ve S.aureus pneumonia bacteria using 0.1g of (both rutile TiO<sub>2</sub>) and (anatase TiO<sub>2</sub>) in separate experiment dark conditions.



**Figure 3.6.a:** Influence of exposure time on the deactivation percentage of  $(1 \times 10^6 \text{ CFU} \text{mL})$  bacteria S. aureus bacteria , catalyst loading 0.1 g from anatase at 28°C and pH=7.4 in dark.



**Figure 3.6.b:** Influence of exposure time on the deactivation percentage of  $(1 \times 10^6 \text{ CFU/mL})$  bacteria S. aureus bacteria , catalyst loading 0.1 g from rutile at28°C and pH=7.4 in dark.

As shown in Fig.(3.5) and (3.6) the deactivation percent was Sharp decline When comparing the two experiences in the dark and under light between (79.29 - 18.6) % for bacteria after 60 min in TiO<sub>2</sub> anatase catalyst .

The percent decreased to ~ 32% for anatase catalysts in dark experiments for G +ve S.aureus after 30 min, whereas the percent decreased to 10% for rutile catalysts in dark experiments for G+ve S.aureus after 30 min and both catalyst not effective at 10 min agents G+ve S.aureus.

Detailed mechanism in dark for  $TiO_2$  anatase work is still not known yet. Most widely accepted mechanism suppose changing in bacterial membrane permeability and dissipation as a result of accumulation and attachment of  $TiO_2$  anatase particles on the bacterial membrane. Bacteria showed big resistance when treated with rutile and the percentage of deactivation was

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very low in both light and dark condition. There for anatase will be used in the flowing studies

## **3.4 TOC results**

4.3.1-TOC results By oxidizing organic carbon supply from deactivated bacteria, the Total Organic Carbon could be perceive and quantified by calculating the amount of oxidized carbon (CO<sub>2</sub>). As shown in Table (4.6) samples from mineralization test ( using 0.1g of TiO<sub>2</sub> anatase ) against  $(1 \times 10^{6} \text{ CFU}\text{mL})$  of S. aureus bacteria were taken after 4 hrs. treatment.

Table 3.4: TOC results for bacterial aqueous mixtures remaining from experiment after 3 hrs. treatment with 0.1 g TiO<sub>2</sub> anatase against  $(1 \times 10^{6} \text{ CFU} \text{mL})$  of S. aureus in light and dark at 28 °C

Sample number	Catalyest appearance	Conditon	Time	T OC mg/L
1,4	TiO <sub>2</sub>	light	4h	28,26
2,5	No –ctalyest	light	4h	46,47
3,6	TiO <sub>2</sub>	dark	4h	39,40
7,8	No –ctalyest	dark	4h	43,42

Table 3.4 show that for S. aureus experiments a clear decrease in organic carbon occurred in light. The waste water treated with  $TiO_2$  showed TOC value 28 mg/L compared to the sample without catalyst which showed 47 mg/L for TOC



Figure 3.7: samples from mineralization experiments (using 0.1g of naked TiO<sub>2</sub> anatase ) against  $(1 \times 10^6 \text{ CFU} \text{mL})$  of S. aureus were taken after 3 h treatment at 28 °C

This difference between the two values confirms that  $TiO_2$  deactivates bacteria, and mineralizes their organic matters under radiation. In dark, the TOC analysis gave almost the same TOC value~ 42 mg\L with and without using TiO<sub>2</sub>. The ability of TiO<sub>2</sub> catalyst in dark to form free radicals is not possible. In dark TiO<sub>2</sub> only deactivates the bacteria with no mineralization.

# 3.5 Impact of concentration of bacteria

Impact of bacterial concentration of S.aureus on bacterial deactivation was studied using 0.1 g naked TiO<sub>2</sub> anatase in treatment process. In Fig. (3.8) 0.1g TiO<sub>2</sub> anatase of was used to treat different concentrations of S.aureus bacteria in separate experiments in light and dark for 60 min at  $28C^{\circ}$ .



Fig 3.8: Impact of increasing the concentrations of S.aureus pneumonia bacteria on the deactivation percentage using 0.1 g of naked  $TiO_2$  in light and dark for 60 min at 28 °C with pH=7.4

Fig. (3.8) shows that when the concentration of S.(aureus) bacteria increases the deactivation percentages decrease. The results show that under light 70.29%, 62.24% and 27.2% of S.aureus respectively, are deactivated in 60 min, from  $1 \times 10^6$  CFU\mL,  $3 \times 10^6$  CFU\mL to  $5 \times 10^6$  CFU\mL solution.

On the other hand, in dark when bacterial concentrations became extremely high the TiO<sub>2</sub> deactivation percent reached only (6.5%) in case of  $5 \times 10^{6}$  CFU\mL of S. aureus.

Table (3.5) illustrate values of QY, TF and deactivation percent values for Different bacterial concentrations of (S. aureus pneumonia bacteria) using 0.1 g of TiO<sub>2</sub> anatase in light for 60 min.

Table 3.5: Values of QY, TF and deactivation percent of increasing the concentrations of S.aureus using 0.1 g of TiO2 anatase in light after 60 min, 28°C.

Concentrations	QY	TN	TF	% deactivated
CFU\mL		min <sup>-1</sup>		bacteria
1×10 <sup>6</sup>	1.664×10 <sup>-44</sup>	9.416×10 <sup>-16</sup>	1.569×10 <sup>-17</sup>	70.29%
3×10 <sup>6</sup>	2.571×10 <sup>-44</sup>	1.48×10 <sup>-15</sup>	2.467×10 <sup>-17</sup>	62.24%
5 ×10 <sup>6</sup>	4.832×10 <sup>-45</sup>	2.78×10 <sup>-16</sup>	4.63×10 <sup>-18</sup>	27.20%

Table (3.5) show that when increasing the concentration of bacteria from  $(1 \times 10^6 \text{ CFU}\text{mL to } 3 \times 10^6 \text{ CFU}\text{mL})$  the value of QY and TF will increase due to the surface of the catalyst is exposed to a greater amount of bacteria, which causes a greater amount of deactivated bacteria.

On other hand when increasing the bacterial concentration more than  $3 \times 10^6$  CFU\mL. The QY, TF will decrease, contrary to what is expected, and this is due to the fact that a large concentration of bacteria performs a process of blocking the sun's rays, which causes a decrease in these values.

# 3.6 Impact of TiO<sub>2</sub> anatase catalyst loading

Lowering and increasing the loading of  $TiO_2$  anatase is necessary for economic reasons. Impacts of this reduction on the deactivation percentage of  $1 \times 10^6$  CFU\mL (S. aureus) in light and dark were studied. Fig. (4.9) shows the results.



**Figure 3.9.a:** Impact of catalyst loading on the deactivation percentage of S. aureus  $(1 \times 10^6 \text{ CFU} \text{mL})$ , after (60) min at 28 °C in light at pH=7.4



**Figure 3.9.b:** Impact of catalyst loading on the deactivation percentage of S. aureus  $(1 \times 10^6 \text{ CFU/mL})$ , after (60) min at 28 °C in dark at pH=7.4

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Fig.(3.9.a) and (3.9.b) shows that in light and dark deactivated percentages were lowered after 60 min to (33.3%, 27.8%) and (11.1%, 7.5%) when 0.05g and 0.025g of TiO<sub>2</sub> were used against S.aureus respectively. By reducing the catalyst weight from 0.1 g to 0.025g catalyst can still remove bacterial.

Table (3.6) shows values of QY, TF and deactivation percent calculated for experiments tested using different  $TiO_2$  anatase loads against  $1 \times 10^6$  CFU\mL of (S. aureus) in light after 60 min.

Table 3.6: Values of QY, TF and % deactivation, for experiments with different TiO<sub>2</sub> anatase loading against  $1 \times 10^6$  CFU\mL of S. aureus in light after 60 min.

loading	QY	TN	TF	% deactivated bacteria
0.1 g	1.664×10 <sup>-44</sup>	9.416×10 <sup>-16</sup>	1.569×10 <sup>-17</sup>	70.29%
0.05 g	9.184×10 <sup>-45</sup>	1.061×10 <sup>-15</sup>	1.768×10 <sup>-17</sup>	33.3%
0.025 g	5.74×10 <sup>-45</sup>	1.326×10 <sup>-15</sup>	2.211×10 <sup>-17</sup>	27.80%

In Tables (3.6) show QY values increase as catalyst weight increases with  $(1.664 \times 10^{-44}, 9.184 \times 10^{-45}, 5.74 \times 10^{-45})$  when (0.1, 0.05, 0.025g) were used against S.aureus bacteria respectively as catalyst loading increased extra photons are used for deactivation.

In other hand table show TF values decreased as catalyst weights increased. This means that relative efficiency of the  $TiO_2$  catalysts decreased with higher loading. This is due to tendency of  $TiO_2$  particles to screen each other from UV light.

## **3.7 Impact of temperature on photo-mineralization reaction**

For S.aureus bacteria, the suitable temperature range for growth is between (7 to 48 °C), with an optimum at 37 °Literature thus confirm that within the range (20-37 °C), the temperature does not have impact on bacterial deactivation. In this work, the Temperature 28 °C was used for technical reasons [51,52]. Due to exposure to light, water temperature stabilizes at 28 C° (slightly higher than room Temperature 25 °C). For this reason , we used the 28 °C [94].

## 3.8 Impact of pH

Impact of pH of was studied on S.aureus bacterial deactivation using 0.1 g  $TiO_2$  anatase in treatment process. In Fig. (4.10), 0.1 g  $TiO_2$  anatase was used with different pH in separate experiments in light and dark for 60 min at 28°C.



**Fig 3.10:** Impact of pH on S.aureu bacteria deactivation percentage using 0.1 g of  $TiO_2$  in light or dark for 60 min at 28 °C at several (pH=5.2,7.4 and 8.8).

The results show the values of deactivated percentage of bacteria are higher in acidic medium than in neutral medium. The neutral medium shows higher degradation than basic medium .This indicates that photodegradation efficiency increased as lower pH value. This is become the cell membrane of bacteria has negative charge and at pH more than (8.8), TiO<sub>2</sub> also has negative charge therefor in basic media repulsive force act as inhibiting factor for reaction between bacteria and TiO<sub>2</sub>. Acidic and neutral media show better photo degradation results.

Values of T.N, Q.Y and degradation percentage were calculated after 60 minutes as shown in Table (4.7).

Table 3.7: Values of QY, TN and % deactivation, for experiments with different pH against  $1 \times 10^6$  CFU\mL of S. aureus in light after 60 min anatase.

pН	% of deactivated Bacteria	TN	QY
5.2	100%	1.06× <b>10<sup>-15</sup></b>	1.83×10 <sup>-44</sup>
7.4	71.92%	9.46×10 <sup>-16</sup>	1.63× <b>10<sup>-44</sup></b>
8.8	88.27%	8.228×10 <sup>-16</sup>	1.42×10 <sup>-44</sup>

The results show that the values of T.N, Q.Y and degradation percentage in acidic medium are higher than in neutral and basic media.

## **3.9 Impact of recovery**

At the end of each round and after sterilizing the samples by autoclave in order to remove of bacteria, new 0.33 ml of bacteria was added to the catalyst was present, light was irradiated for 1 hour. Dark experiments were also conducted for them.



**Fig 3.11.a:** 0.1g of TiO<sub>2</sub> anatase were reused for 3rd time against of S. aureus  $(1 \times 10^6 \text{ CFU/mL})$ , for 60 min. at 28 °C dark condition.



**Fig 3.11.b**: 0.1g of TiO<sub>2</sub> anatase were reused for 3rd time against of S. aureus ( $1 \times 10^6$  CFU\mL), for 60 min. at 28 °C at light condition.

Fig.(4.11.a) and (4.11.b) shows possibility to reuse the catalyst for third time in light and in dark.

in light condition deactivation percent lowering from 100% to 32% in second round and lowering to 25.9% in third round, in dark condition deactivation percent lowering from 43.28 % to19.6% in second round and lowering to 6.11% in third round

noting the decrease in the efficiency and ability of the catalyst to gradually inactivate the bacteria whenever it is reused.

## **3.10 Conclusion and Recommendations**

The results show that  $TiO_2$  anatase can be effective catalyst to deactivate G +ve S. aureus bacteria in both dark and light experiments. Under light, mineralization of bacteria occurs for S. aureus (43.47%) for the first time  $TiO_2$  catalysts can also be recovered and reused for second time.

The results show the potential of using  $TiO_2$  catalyst system in future waste water disinfection. For future work, we suggest applying the  $TiO_2$  catalyst against other types of bacteria and other species such as viruses and algae Applying the  $TiO_2$  catalyst on sewage water, in cooperation with the Water Authority is recommended. The study should involve both organic contaminants and microorganisms.

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جامعه النجاح الوطنية كلية الدراسات العليا

## تقييم الدور الحفزي الضوئي لدقائق اكسيد التيتانيوم في التحطيم الكامل للبكتيريا ومحتوياتها العضوية في الماء

إعداد رائد رسمي حسن علي

إشراف أ. د. حکمت هلال د. عاهد زيود

قدمت هذه الأطروحة استكمالا لمتطلبات الحصول على درجة الماجستير في برنامج الكيمياء، من كلية الدراسات العليا، في جامعة النجاح الوطنية، نابلس – فلسطين. تقييم الدور الحفزي الضوئي لدقائق اكسيد التيتانيوم في التحطيم الكامل للبكتيريا ومحتوياتها العضوية في الماء إعداد رائد رسمي حسن على إشراف د. حكمت هلال د. عاهد زيود الملخص

تتقية المياه وتطهيرها من الملوثات المتعددة أصبح ضرورة لحياة كريمة للفرد، حيث يتم إتباع عدة استراتيجيات وطرق مختلفة في دول عدة من أجل تنقية المياه من هذه الملوثات، ان أفضل الطرق هي الطرق الآمنة على صحة الإنسان وذات الكفاءة العالية والتكلفة المنخفضة.

إن التحطيم الضوئي للبكتيريا يعتبر من أكثر الطرق كفاءة و جذبا لاهتمام الباحثين في هذا المجال حيث يتم تحفيز أشباه الموصلات باستخدام الضوء والذي يقوم بدوره بالتحطيم الضوئي للملوثات البيولوجية والكيميائية المتواجدة في الماء حيث أصبحت حبيبات أكسيد التيتانيوم النانوية (TiO<sub>2</sub>) تستخدم على نطاق واسع لكثير من الملوثات العضوية في الماء.

إن حبيبان ثاني اكسيد التيتانيوم النانوية (TiO<sub>2</sub>) التي تمتلك فجوة طاقة (Band gap) واسعة من 2.5 الى 3 الكترون فولت تحتاج إلى طول موجة يتواجد في منطقة الأشعة فوق البنفسجية حيث يقتصر النشاط التحفيزي على أطوال موجات أقصر تقع فهذه المنطقة.

في هذا العمل تم تعقيم المياه عن طريق التحطيم الكامل لنوع البكتيريا إيجابية الغرام (Staphylococcus aureus) مع مكوناتها العضوية وذلك باستخدام جزيئات ثاني أكسيد (Staphylococcus aureus) مع مكوناتها العضوية وذلك باستخدام جزيئات ثاني أكسيد التيتانيوم النانوية (TiO<sub>2</sub> anatase) حيث لوحظ انه بعد 60 دقيقة تم التخلص من البكتيريا بنسبة 70,29 وبعد 4 ساعات لوحظ انخفاض كبير في المكونات العضوية الكربونية بنسبة 55% ولقد تمت دراسة أثر بعض العوامل على فعالية الحفاز وسير تفاعل التحطيم الضوئي مثل درجة الحرارة والفترة الزمنية للتعريض الضوئي ودرجة الحموضة وتركيز كل من الحفاز والملوثات هذا وقد تبين إن الحفاز يمكن إعادة استخدامه اكثر من مرة.