Molecular and Serologic Approaches for Diagnosis of *Mycoplasma* pneumoniae Infections in Children and Typing of the Pathogen.

Motasem Almasri^{1,2}, Efimia Souliou², Ana Papa², Nikos Mallisiobas², Euthoxia Diza², Antoniadis²

Introduction

M. pneumoniae is a common cause of upper and lower respiratory tract infections of variable severity (4). Infections of this pathogen occur worldwide throughout the year (7), although they are best documented as a significant cause of respiratory disease in temperate climates (3). The incidence of M. pneumoniae is greatest among school-age children and declines after adolescence (1). However, M. pneumoniae may occur endemically and occasionally epidemically in older persons, as well as in children under five years of age (2). The clinical presentation of patients with M. pneumoniae infection is not significantly different from that of patients with infections caused by other bacteria or viruses, so diagnosis of M. pneumoniae infection relies primarily on special laboratory testing (6). The lack of rapid and accurate diagnostic laboratory tests to detect M. pneumoniae directly or the serologic response it elicits has hampered understanding of the epidemiology and contributed to the unawareness of the potential clinical significance of this common pathogen (9). A specific diagnosis is important, because b-lactam antibiotics treatment of an infection due to M. pneumoniae is ineffective, whereas the use of other types of antibiotics, such as macrolides, can markedly reduce the duration of the illness (5). The clinical isolates of M. pneumoniae have been shown to exhibit some nucleotide sequence variation in the P1 gene.

Accordingly, the *M. pneumoniae* strains were divided into two groups, I and II (8).

Materials and Methods

The study included 225 children hospitalized for respiratory tract infections during a13-month period (1 May 2003 to 1 June 2004), in the Department of Pediatrics of AHEPA Hospital-Aristotle University of Thessaloniki. The children were 2 months to 14 years old. From each patient a throat swab and a serum specimen were taken upon admission. A second serum specimen was taken 5-20 days later. Throat swab specimens from these children were examined for the detection of *M. pneumoniae* by: Methylene blue-glucose diphasic medium, EIA-antigen detection and PCR. Both of the first and second serum specimens from these patients were examined for the presence of antibodies against *M. pneumoniae* by complement fixation test (CFT), Indirect Immunofluorescence assay-IFA (IgM and IgG) and Enzyme Linked Immunoassay-ELISA (IgM, IgG, and IgA). A region in the P1 gene (1110 bp) was sequenced for typing of *M. pneumoniae* strains. Using CLUSTAL W program the DNA sequences of the study strains were compared to each other as well as to DNA sequences of P1 gene of reference strains.

Results

A confirmed laboratory result suggestive of current *M. pneumoniae* infection was obtained for a total of 23 children (10.2%) of the patients. In these 23 children, culture, antigen detection test and PCR were positive in 4 cases (sensitivity 17.4%), 1 (4.4%) and 17 (73.9%), respectively. In the acute phase first serum specimens CFT, IgM-IFA, IgM-ELISA and IgA-ELISA were positive in 7 (sensitivity 30.4%), 11 (47.8%), 16 (69.6%) and 2 (8.7%), respectively. The specificities of these methods were found as follows: culture 100%, antigen detection test 100%, PCR 96.5% and for serologic methods: CFT 97.5%, IgM-IFA 91.6% and IgM-ELISA 98%. The combination of PCR and IgM-capture ELISA provides highly sensitive and accurate tool for the rapid diagnosis of *M. pneumoniae* infection in children. In

¹ Department of Biology and Biotechnology, An Najah National University, Nablus, Palestine.

²A' Department of Microbiology, School of Medicine, Aristotle University of Thessalonoki, Greece.

Greece both types (I and II) of *M. pneumoniae* existed and type I was predominant in 2003. There were a considerable differences in the nucleotide sequence between the 2 types of *M. pneumoniae* most of which were reflected in the amino acid sequence of the P1 protein.

References

- Block S., Hedrick J., Hammerschlag M.R., Cassell G.H., and Craft J.C. (1995). Mycoplasma pneumoniae and Chlamydia pneumoniae in pediatric community-acquired pneumonia: comparative efficacy and safety of clarithromycin vs. erythromycin ethylsuccinate. Pediatr Infect Dis J, 14:471–477
- Dorigo-Zetsma J.W., Wilbrink B., van der Nat H., Bartelds A.I.M., Heijnen M.A., and Dankert J. (2001). Results of molecular detection of Mycoplasma pneumoniae among patients with acute respiratory infection and in their household contacts reveal children as human reservoirs. J Infect Dis, 183:675–678.
- Fleming P.C., Kreiger E., Turner J.A.P. et al. (1967). Febrile mucocutaneous syndrome with respiratory involvement, associated with isolation of Mycoplasma pneumoniae. Can Med Assoc J, 97:1458
- Foy H.M. Mycoplasma pneumoniae: current perspectives. (1999). Clin Infect Dis, 28:237.
- Hammerschlag M.R. Atypical pneumonias in children. (1995). Adv Pediatr Infect Dis,10:1–39.
- Lieberman D., Schlaeffer F., Lieberman D., Horowitz S., Horovitz O., and Porath A. (1996). Mycoplasma pneumoniae community-acquired pneumonia: a review of 101 hospitalized adult patients. Respiration, 63:261–266.
- Murray P.R., Rosenthal K.S., Kobayashi G.S., and Pfaller M.A.(1998). Medical Microbiology. 3rd edn. New York: Mosby, 347-351.
- Su C.J., Dallo S.F., Chavoya A., and Baseman JB. (1993). Possible origin of sequence divergence in the P1 adhesin gene of Mycoplasma pneumoniae. Infect Immun,61:816–822.
- Waites K.B. (2003). New Concepts of Mycoplasma pneumoniae Infections in Children. Pediat Pulmonology, 36:267–278.